

# Modelling and surveillance of infectious diseases - or why there is an in SARS

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useR!2008  
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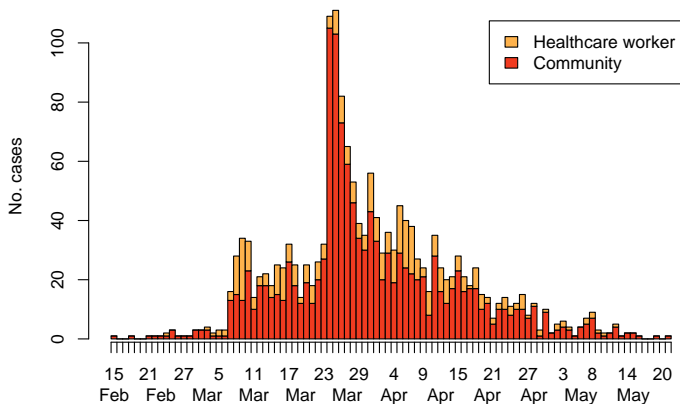
# Motivation

- How can R assist in understanding and controlling infectious diseases – be it in human, plant or veterinary epidemiology.
- Two R packages exist:
  - ① `RLadyBug` contains a set of functions for the simulation and parameter estimation in spatially heterogeneous SIR models.
  - ② `surveillance` contains algorithms for the detection of aberrations in time series of counts arising from routine public health surveillance
- This talk intends to give an overview of using R for especially (1) – deeper mathematical details are suspended to the lunch break

```
> library("RLadyBug")
```

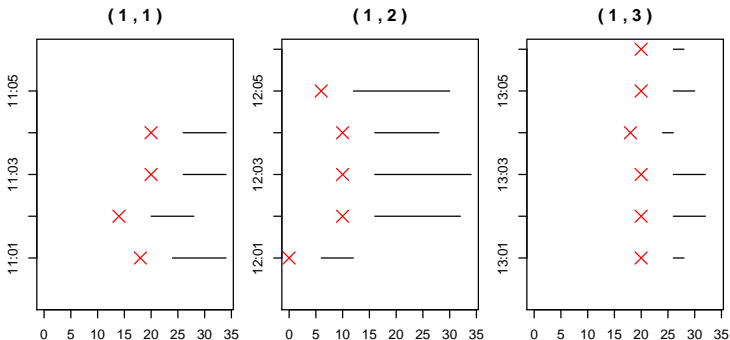
## Applications (1): SARS in Hong Kong 2003

- Daily number of new cases of the severe acute respiratory syndrome (SARS) in Hong Kong (Anonymous, 2003)
- Epidemic curve created with package epitools (Aragon, 2007).



## Applications (2): CSF Transmission Experiment

- Experiment by Laevens et al. (1999) with classical swine fever (CSF) using  $\mathbf{S}(0) = (5, 5, 6)$  and  $\mathbf{E} = (0, 1, 0)$ .
- Event history of each pig with inoculation as origin

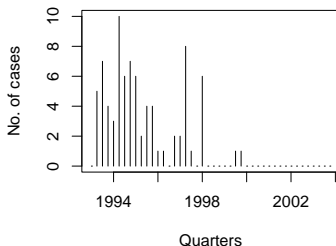


```
> data("laevens")
> plot(laevens, type = individual ~ time | position)
```

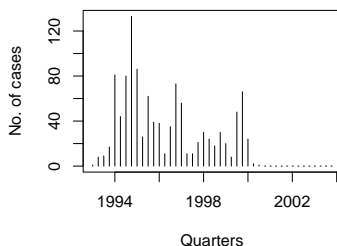
## Applications (3) – CSF surveillance

- Classical swine fever (CSF) in Brandenburg (BB) and Mecklenburg-Western Pomerania (MP), Germany
- Total of 81 infected farms out of 3290 during 1993-2004

**CSF in domestic pig farms in MPBB**



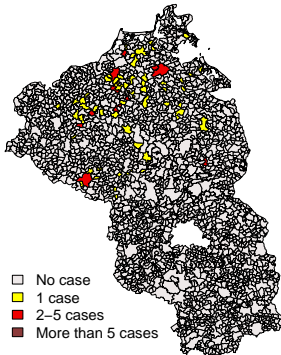
**CSF among wild boars in MPBB**



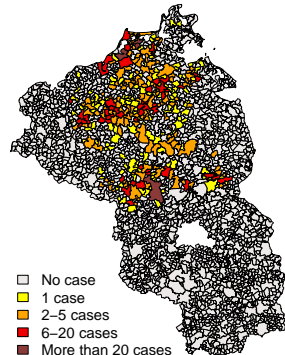
- Interest in investigating the connection between the CSF incidence among domestic pigs and wild boars

# Applications (4) - Spatial incidence of CSF in MPBB

**Domestic pigs in MPBB**



**Wild boars in MPBB**



## Stochastic epidemic models (1)

- SEIR model: A closed population of  $n + m$  individuals divided into susceptible, exposed, infectious, and recovered
- $S(0) = n$ ,  $E(0) = m$ ,  $I(0) = 0$  and  $R(0) = 0$
- At time  $t$ , an individual  $j$  meets infectious at rate

$$\lambda_j(t|\mathcal{H}_t) = \sum_{i=1}^{n+m} \mathbb{1}_{i \in \text{Infectious}(t)} \cdot f(i, j),$$

where  $f(\cdot) \geq 0$  is a function of the distance between  $i$  and  $j$

- If a susceptible meets an infected, it becomes exposed

## Stochastic epidemic models (2) – Distance kernels

- ① Homogeneous model:  $\forall i, j : f(i, j) = \beta > 0$  and hence

$$\lambda_j(t|\mathcal{H}_t) = \beta I(t)$$

- ② Heterogeneous model: The population is made up of  $k$  units arranged on a grid in space. For  $j$  in unit  $u_j$ :

$$\lambda_j(t|\mathcal{H}_t) = \beta I_{u_j}(t) + \beta_\eta \sum_{u \in N(u_j)} I_u(t)$$

- ③ Heterogeneous model, where individuals have locations in  $\mathbb{R}^2$  and  $f(i, j)$  is a function on the Euclidean distance  $\text{dist}(i, j)$



## SARS in Hong Kong 2003

- Assuming a constant incubation time of 6.4 days and a constant recovery time of 34 days as suggested by the mean of the distributions in Donnelly et al. (2003) we obtain

```
> data("hksars")  
> print(m1 <- seir(hksars, hksars.opts.ml))
```

```
Calling LadyBug (monitor ladybug.system.out/err for progress)...  
...  
Parameter Estimations:  
Parameter:  
      beta  
4.3984e-09  
...
```

- Basic reproduction number  $R_0 = R_0(m1, hksars) = 1.0012$ .

## CSF Transmission Experiment (1)

- Exposure times are not observed, instead of imposing we assume  $T_E \sim \mathcal{G}a(\delta_E, \gamma_E)$  and  $T_I \sim \mathcal{G}a(\delta_I, \gamma_I)$
- A Bayesian setting with MCMC is used for parameter inference

```
> print(m2 <- seir(laevens, laevens.opts.mcmc))
```

An object of class `LBInferenceMCMC`

Parameter Estimations (posterior mean from 2500 samples):

Parameter:

beta	betaN	gammaE	deltaE	gammaI	deltaI
0.03706	0.02837	56.82000	9.37400	2.16200	0.25640

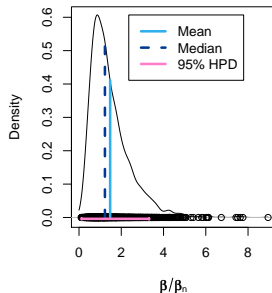
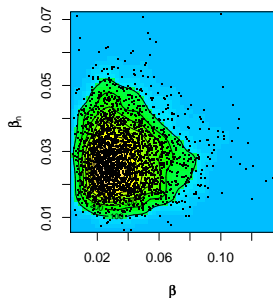
StandardErrors (posterior std.dev. from 2500 samples):

beta	betaN	gammaE	deltaE	gammaI	deltaI
0.018500	0.009481	45.510000	7.761000	0.738100	0.097760

## CSF Transmission Experiment (2)

- MCMC output can be further analysed by e.g. coda package
- Posterior density of  $\beta/\beta_n$  and  $R_0$

```
> plot(m2, which = "betabetaN")
> quantile(R0(m2, laevens), c(0.025, 0.5, 0.975))
```



```
2.5%    50%    97.5%
0.3245 0.6370 1.2426
```

## CSF surveillance (1)

- CSF surveillance data consists of multiple outbreaks
- SIR Extension: Risk of infection consists of two components
  - **endemic component**: Time to infection from external sources modelled by a Cox model
  - **epidemic component**: Similar to heterogeneous SIR model with distance weighting of infectives
- Rate of infection has the following form

$$\lambda_j(t|\mathcal{H}_t) = \exp(h_0(t) + \mathbf{z}_j(t)' \boldsymbol{\alpha}) + \sum_{i=1}^{n+m} \mathbb{1}_{i \in \text{Infectious}(t)} \cdot f(i, j)$$

- When using a linear basis expansion of  $f(i, j)$  this rate is similar to the conditional intensity of an additive-multiplicative hazard model from survival analysis

## CSF surveillance (2)

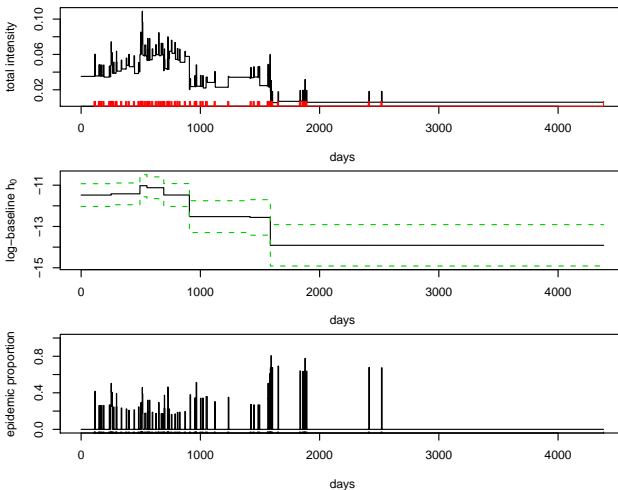
- Endemic component: piecewise exponential baseline and time varying covariates boars and vaccination area
- Epidemic component:  $f(i, j) = \beta > 0$
- Inference using penalized loglikelihood with a model syntax similar to the `timereg` package (Scheike, 2006)

```
> m3 <- spatialSIR(Surv(start, stop, event) ~ fconst +  
+   cox(boar) + cox(vacc), data = mpbb.evHist, ...)  
> coef(m3)[c("fconst", "cox(boar)", "cox(vacc)")]  
> diag(vcov(m3))[c("fconst", "cox(boar)", "cox(vacc)")]
```

```
  fconst cox(boar) cox(vacc)  
3.814e-06 2.108e+00 1.261e+00  
  
  fconst cox(boar) cox(vacc)  
7.371e-12 9.263e-02 1.729e-01
```

## CSF surveillance (3)

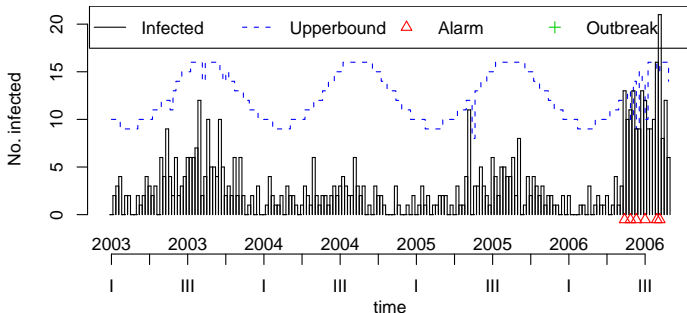
- Plot of the total intensity  $\sum_{j=1}^{n+m} \lambda_j(t|\mathcal{H}_t)$ , the log baseline hazard  $h_0(t)$  (with a 95% CI) and the epidemic proportion



# The surveillance package (1)

```
> library("surveillance")  
> data("shadar")  
> control = list(range = 105:295, ret = "cases", alpha = 0,  
+   c.ARL = 5)  
> plot(algo.glrnb(shadar, control = control))
```

## Analysis of shadar using glrpois: intercept



## The surveillance package (2)

- Surveillance algorithms:
  - `cdc` – Stroup et al. (1989)
  - `farrington` – Farrington et al. (1996)
  - `cusum` – Rossi et al. (1999)
  - `roya` – Rogerson and Yamada (2004) (Experimental)
  - `lr` and `glr` – H. and Paul (2008)
- Comparison of surveillance algorithms using sensitivity, specificity and its variants in simulations
- Surveillance time series models:
  - `hhh` - Held et al. (2005); Paul et al. (2008)
  - `twins` - Held et al. (2006) (Experimental)



## Discussion

- First steps towards R functionality for infectious disease modelling. More complex and realistic models imaginable.
- Packages contain many additional visualization and simulation procedures (Sellke construction, Ogata's modified thinning)
- Combining database, R, Sweave/odfWeave and LaTeX/OpenOffice allows for easy generation of daily bulletins or reports

```
> motd
```

### Message of the day

Packages are on CRAN. Starting points are H. (2007); H. and Feldmann (2007). Maybe they are of help. If adaptation is needed for your problem let me know.

# Acknowledgements

## Persons:

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